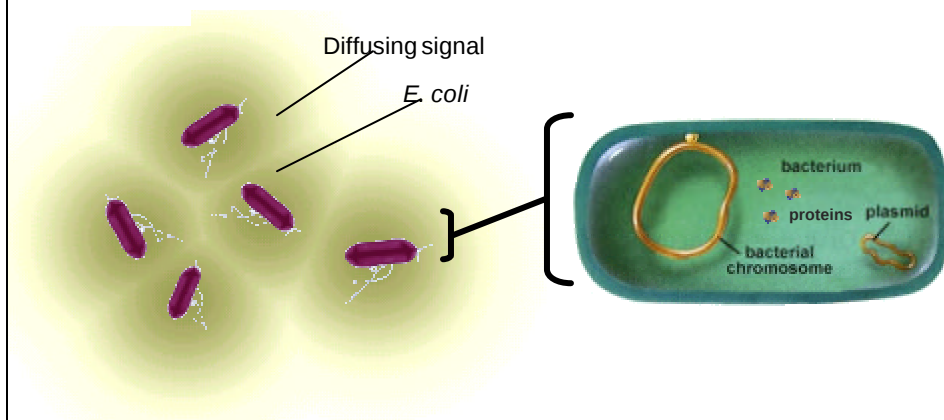


Cell-Cell Communications

Ron Weiss
Department of Electrical Engineering
Princeton University

Computing Beyond Silicon Summer School, Caltech, Summer 2002

Programming Cell Communities

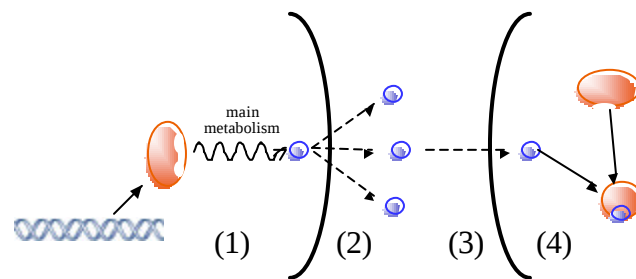


Program cells to perform various tasks using:

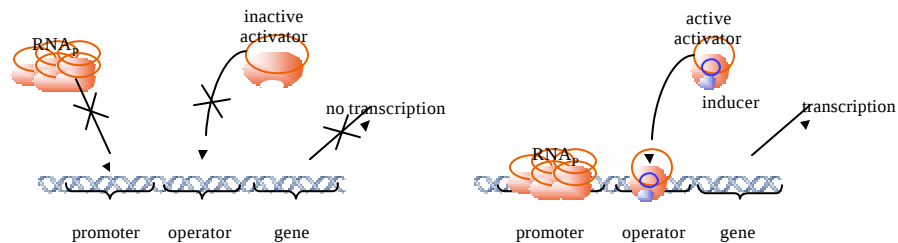
- Intra-cellular circuits
 - Digital & analog components
- Inter-cellular communication
 - Control outgoing signals, process incoming signals

Intercellular Communications

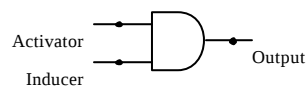
- Certain inducers useful for communications:
 1. A cell produces inducer
 2. Inducer diffuses outside the cell
 3. Inducer enters another cell
 4. Inducer interacts with repressor/activator → change signal



The Intercellular AND Gate



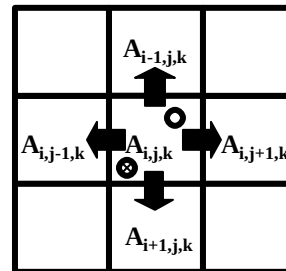
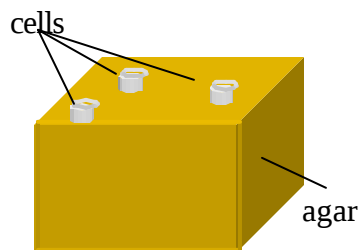
- Inducers can activate activators:
 - VAI (3-N-oxohexanoyl-L-Homoserine lacton) → luxR
- Use as a logical AND gate:



Activator	Inducer	Output
0	0	0
0	1	0
1	0	0
1	1	1

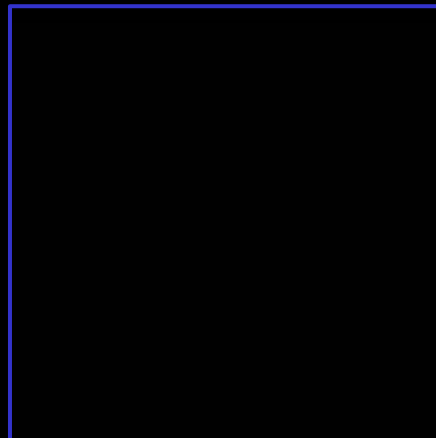
Communications Simulator

- Cells sit on 3D agar grid
- Model genetic networks in cells (ODE, stochastic)
- ODE diffusion model with reflective boundaries



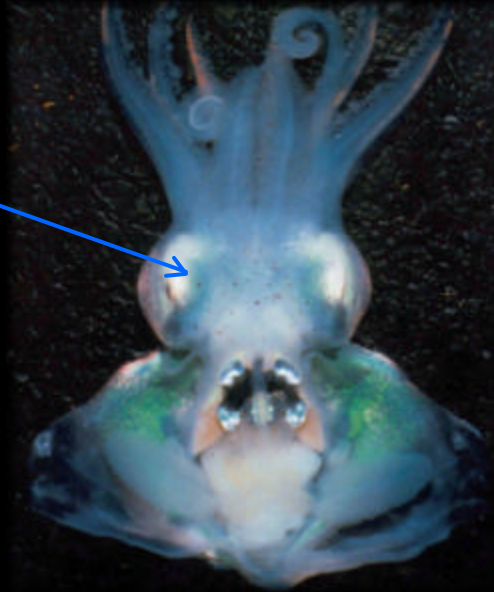
3D diffusion:
$$dA_{i,j,k}/dt = k_{\text{diff}} (A_{i,j-1,k} + A_{i,j+1,k} + A_{i-1,j,k} + A_{i+1,j,k} + A_{i,j,k+1} + A_{i,j,k-1} - 6A_{i,j,k})$$

Two Cell Simulation



Eupryma scolopes

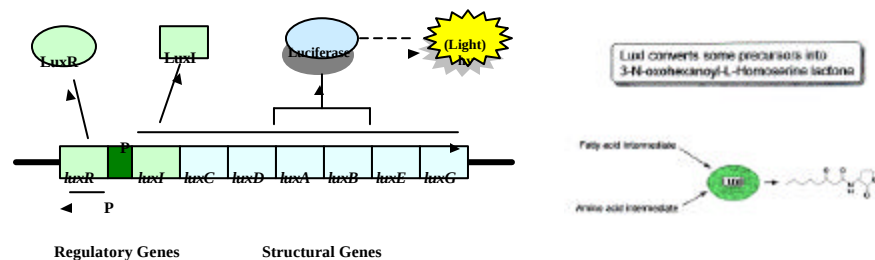
Light organ



Quorum Sensing

- Cell density dependent gene expression

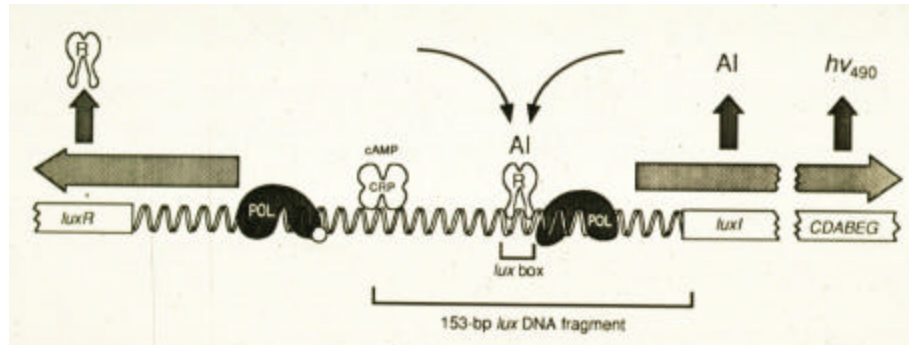
Example: *Vibrio fischeri* [density dependent bioluminescence]



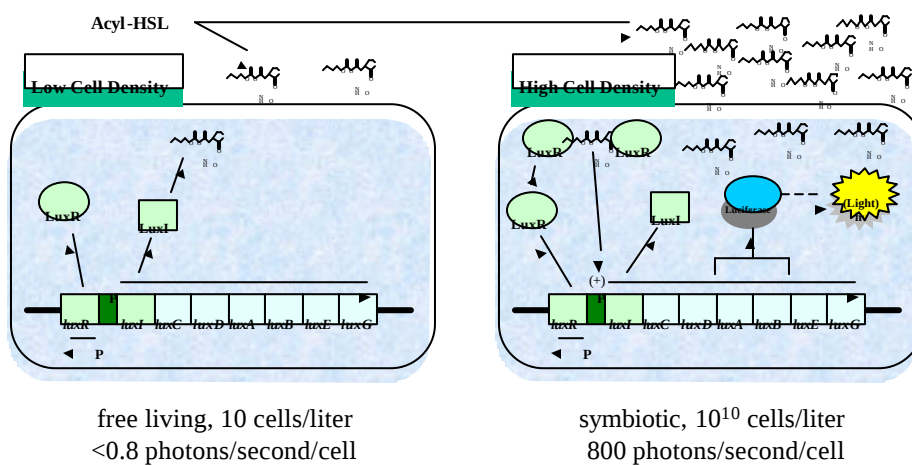
The *lux* Operon

LuxI metabolism
→ autoinducer (VAI)

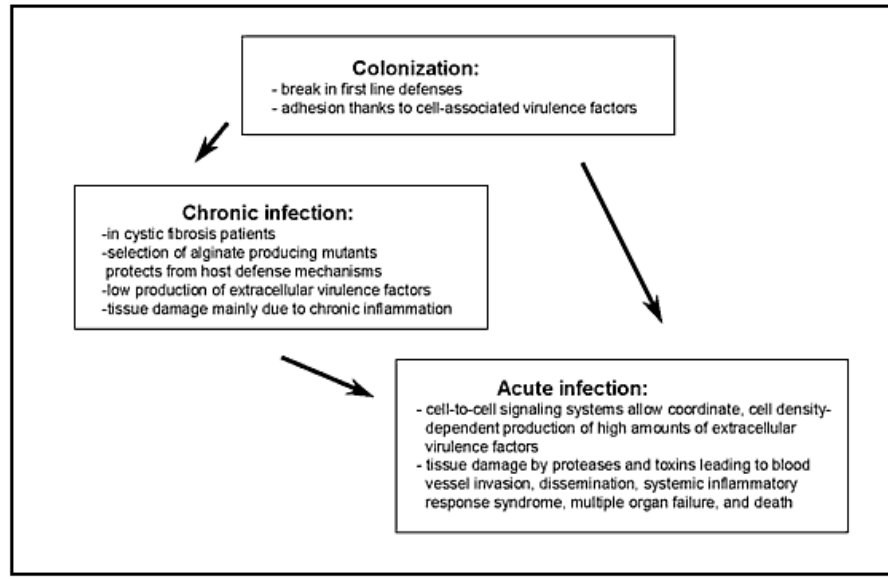
The lux box



Low and High Cell Densities

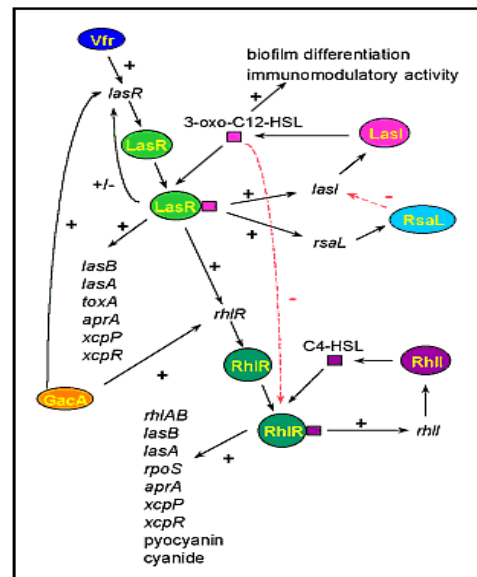


P. Aeruginosa



P. Aeruginosa

- Two autoinducer systems regulate virulence/biofilm formation
- Secrete virulence factors when population high enough to overcome host defenses

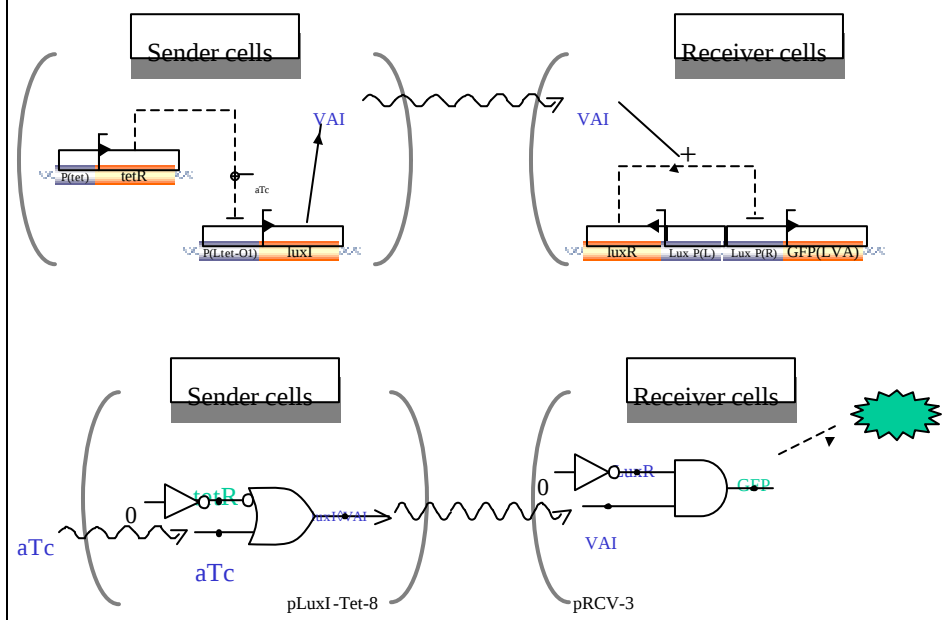


Sources for a Library of Signals

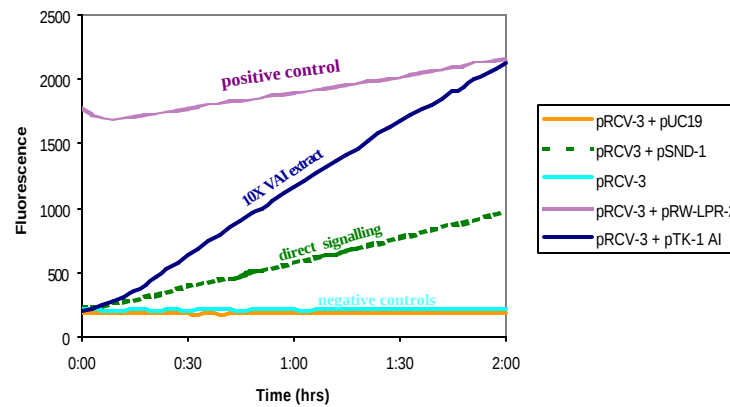
N-acyl-L-Homoserine Lactone Autoinducers in Bacteria

Species	Relation to Host	Regulate Production of	I Gene	R Gene
<i>Vibrio fischeri</i>	marine symbiont	Bioluminescence	<i>luxI</i>	<i>luxR</i>
<i>Vibrio harveyi</i>	marine symbiont	Bioluminescence	<i>luxL,M</i>	<i>luxN,P,Q</i>
<i>Pseudomonas aeruginosa</i>	Human pathogen	Virulence factors	<i>lasI</i>	<i>lasR</i>
		Rhamnolipids	<i>rhlI</i>	<i>rhlR</i>
<i>Yersinia enterocolitica</i>	Human pathogen	?	<i>yenI</i>	<i>yenR</i>
<i>Chromobacterium violaceum</i>	Human pathogen	Violaceum production Hemolysin Exoprotease	<i>cvrI</i>	<i>cvrR</i>
<i>Enterobacter agglomerans</i>	Human pathogen	?	<i>eaqI</i>	?
<i>Agrobacterium tumefaciens</i>	Plant pathogen	Ti plasmid conjugation	<i>traI</i>	<i>traR</i>
<i>Erwinia caratovora</i>	Plant pathogen	Virulence factors Carbapenem production	<i>expI</i>	<i>expR</i>
<i>Erwinia stewartii</i>	Plant pathogen	Extracellular Capsule	<i>esaI</i>	<i>esaR</i>
<i>Rhizobium leguminosarum</i>	Plant symbiont	Rhizome interactions	<i>rhiI</i>	<i>rhiR</i>
<i>Pseudomonas aureofaciens</i>	Plant beneficial	Phenazine production	<i>phzI</i>	<i>phzR</i>

Cell-Cell Communication Circuits

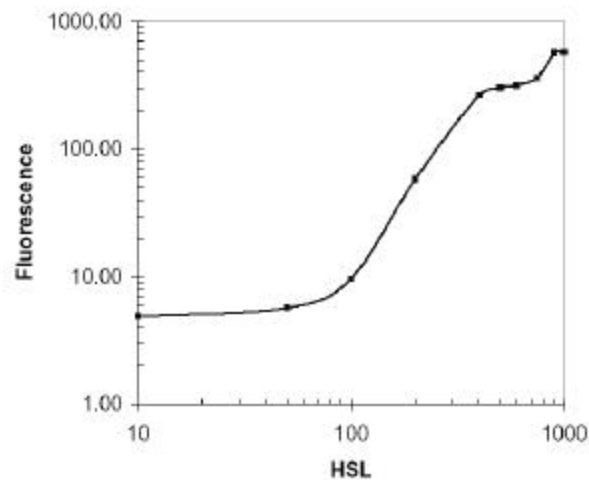


Time-Series Response to Signal



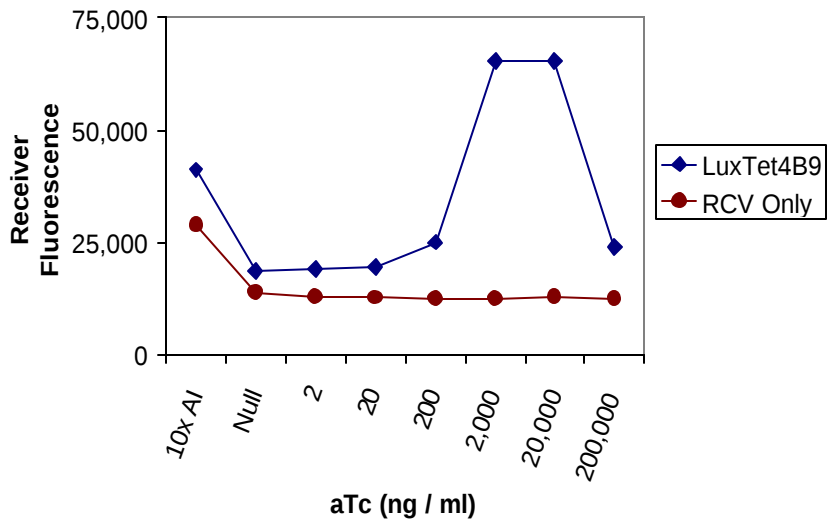
Fluorescence response of receiver (pRCV-3)

Characterizing the Receiver

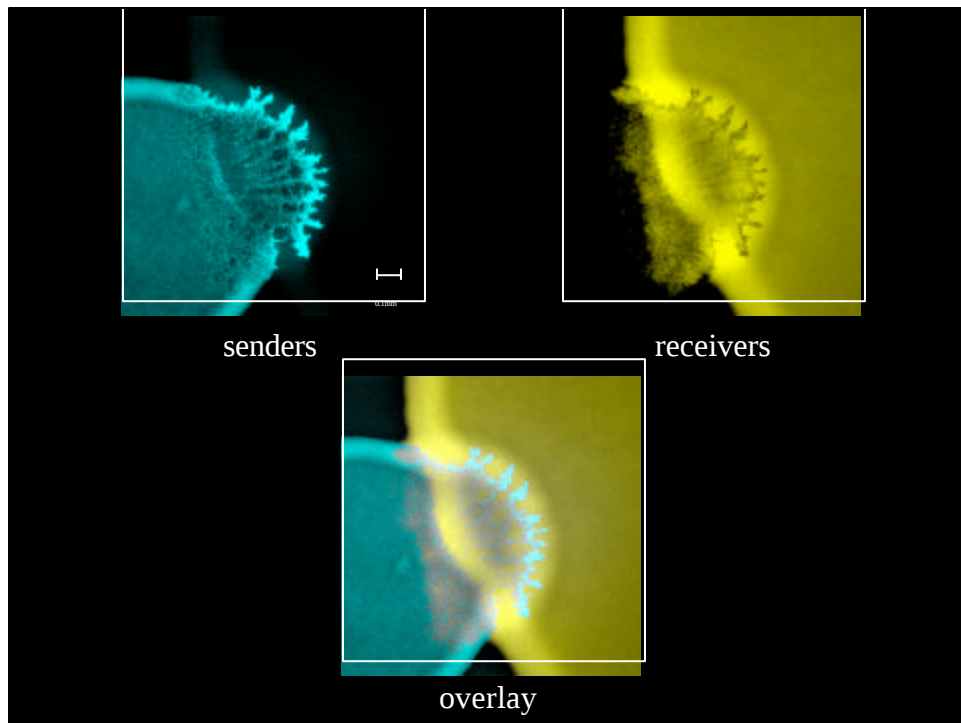


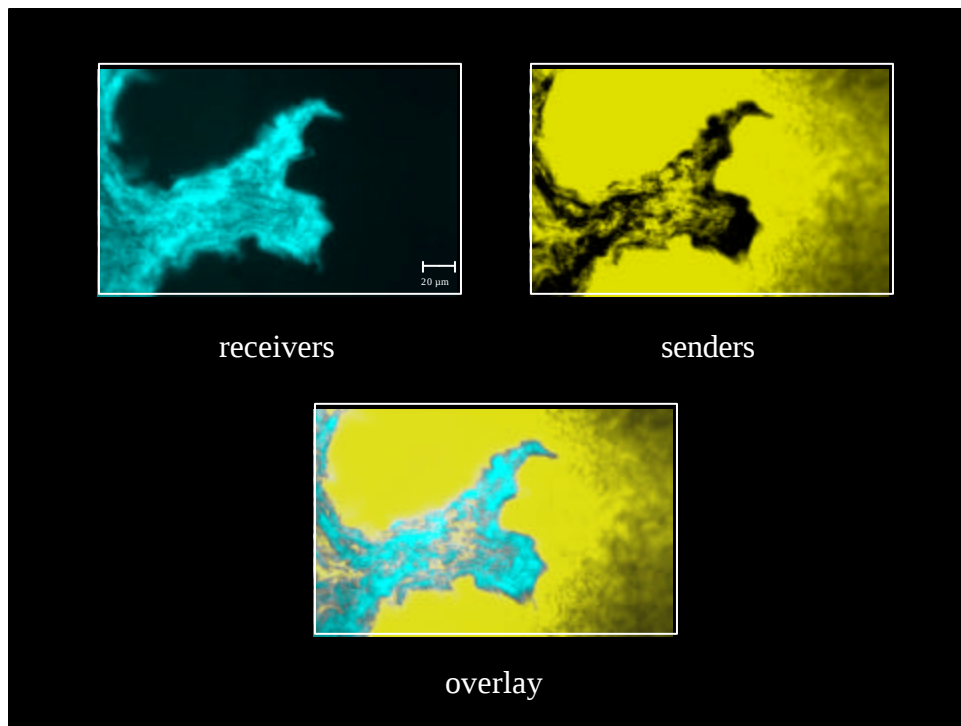
Response of receiver to different levels of VAI extract

Controlling the Sender's Signal Strength



Dose response of receiver cells to aTc induction of senders





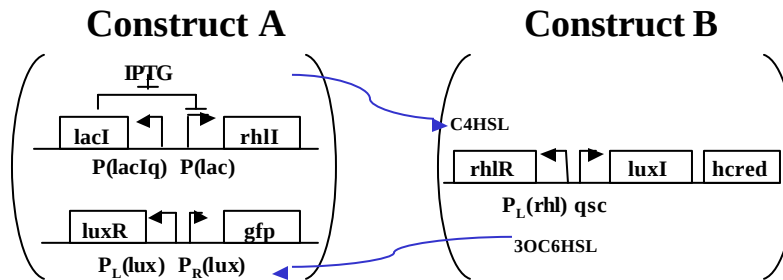
Engineered Microbial Communication

Ron Weiss
Tom Knight
Nick Papadakis

February, 2001

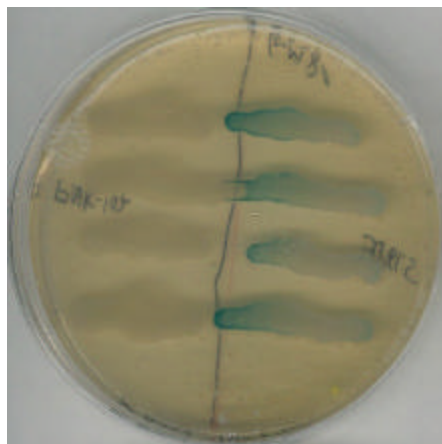
Bi-Directional Communication

[Karig, Weiss]



- Explore substrate properties
 - Crosstalk
 - Time scale/delay
 - Signal strength
- Create constructs useful in later systems

Demonstrating rhII Communications

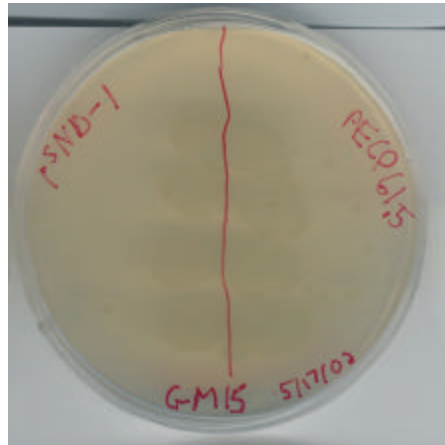


senders

receivers

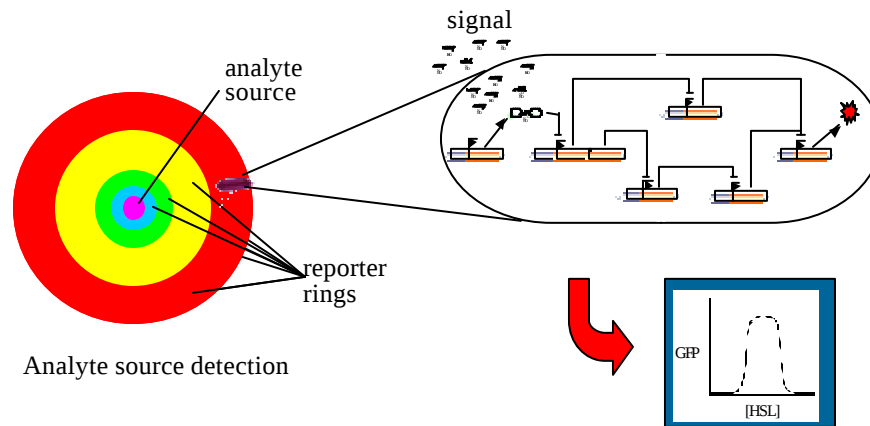
Testing Crosstalk

Does 3OC6HSL bind RhlR to activate transcription?

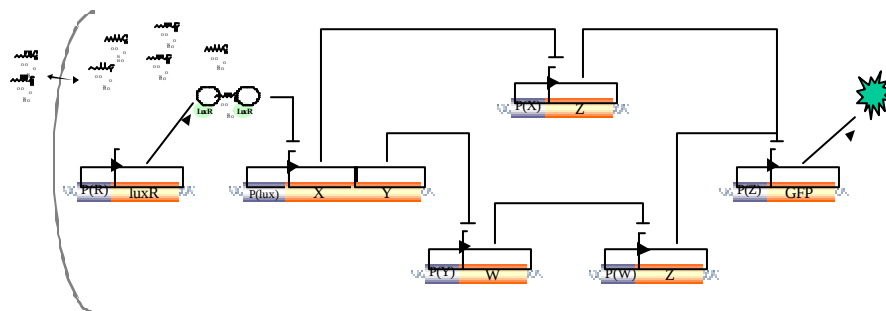


Signal Processing / Analog Circuits

Detecting Chemical Gradients

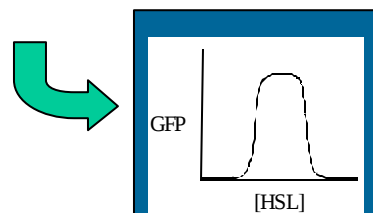


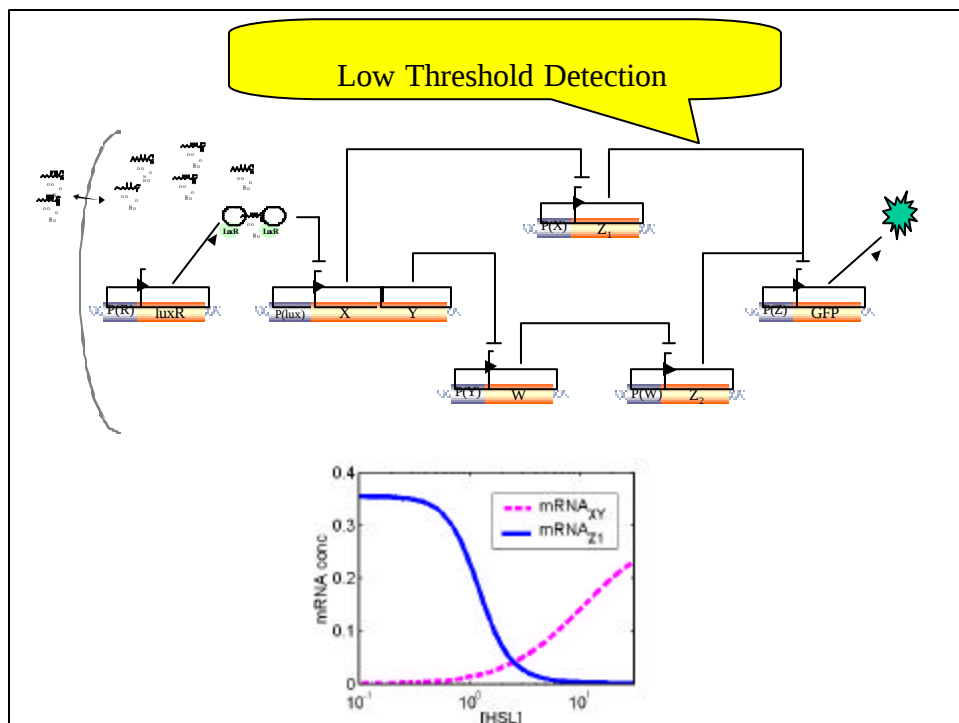
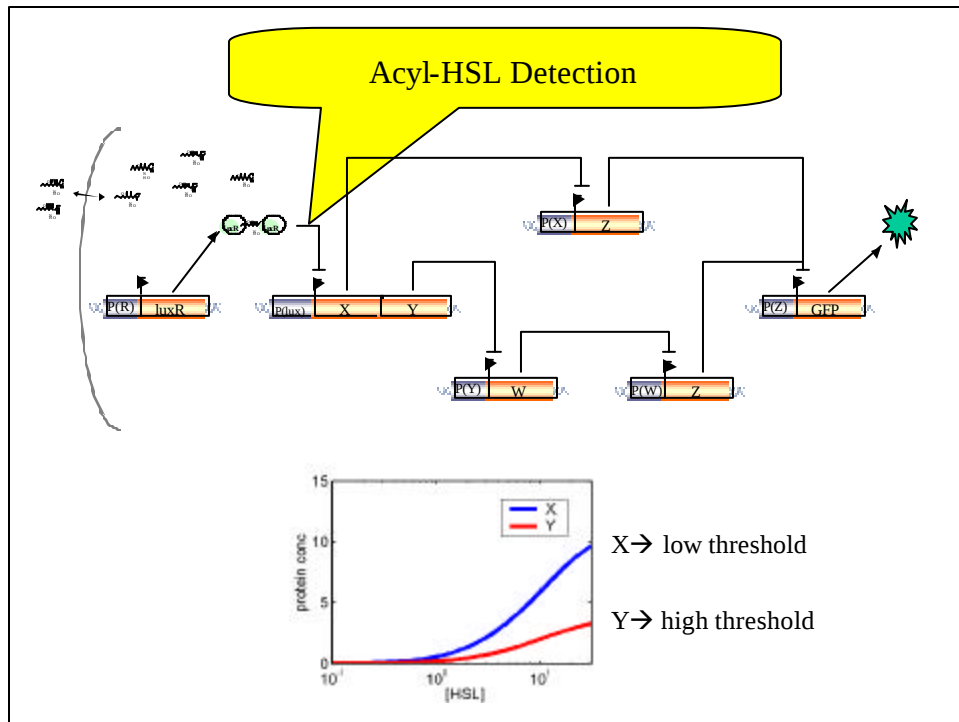
Circuit Components

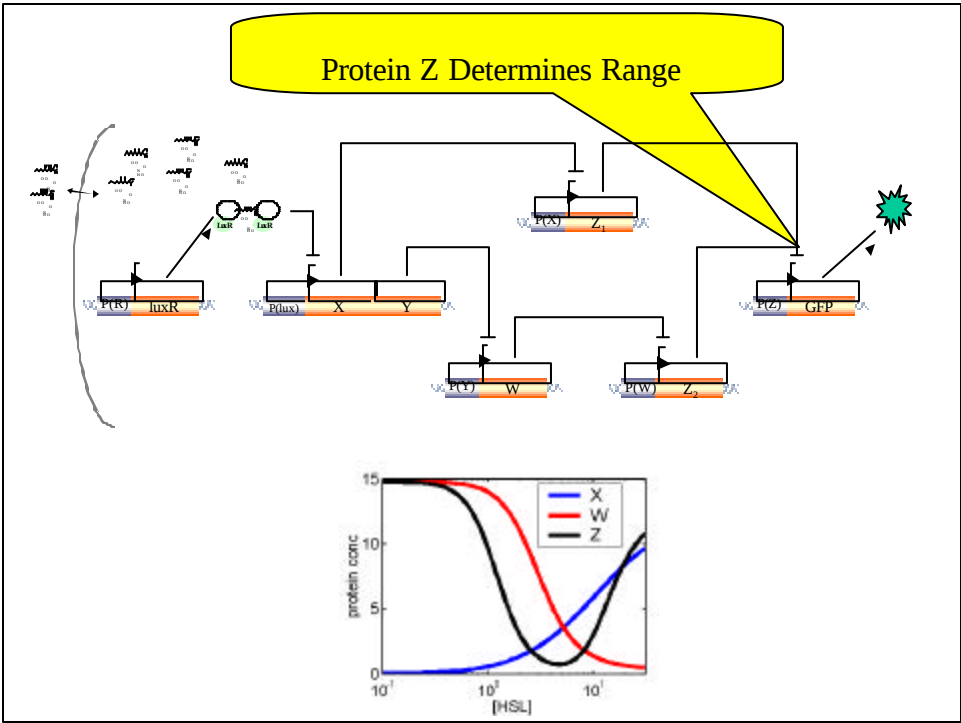
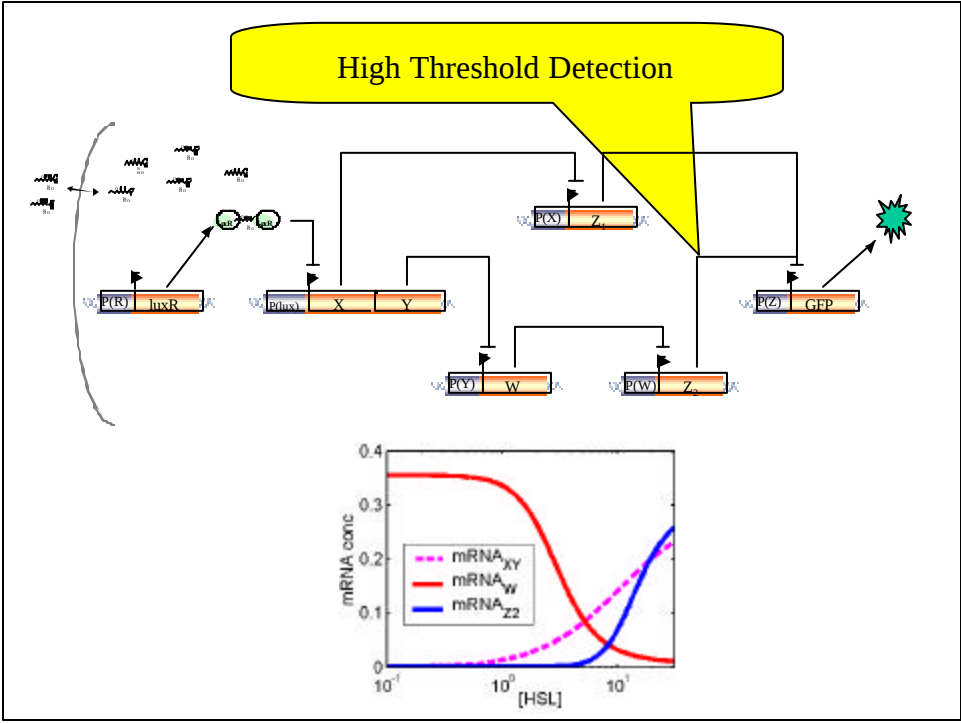


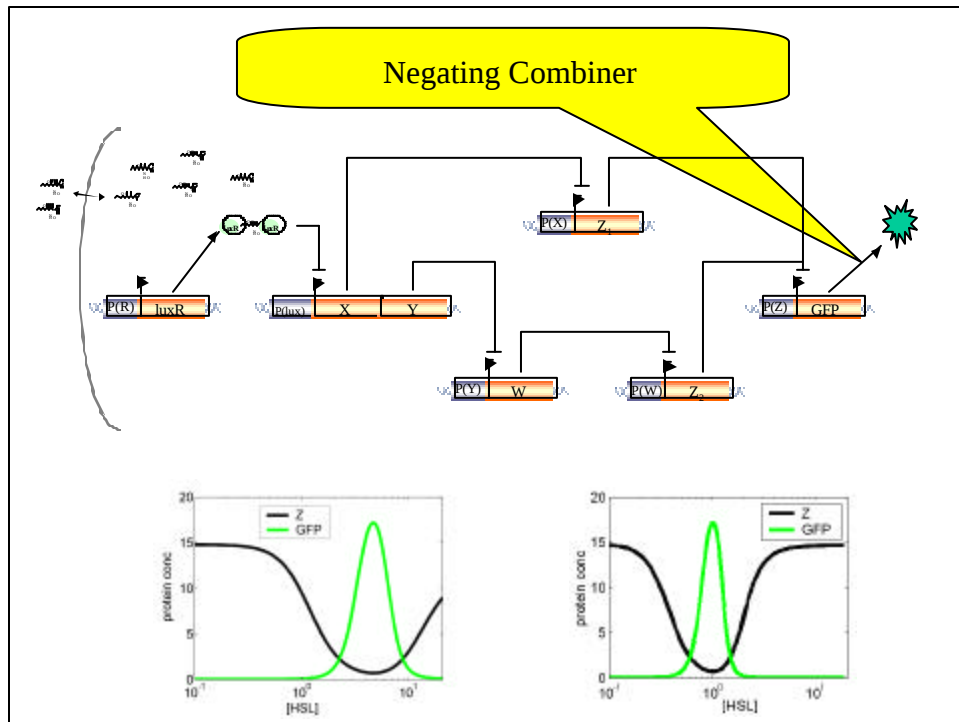
Components:

1. Acyl-HSL detect
2. Low threshold
3. High threshold
4. Negating combiner



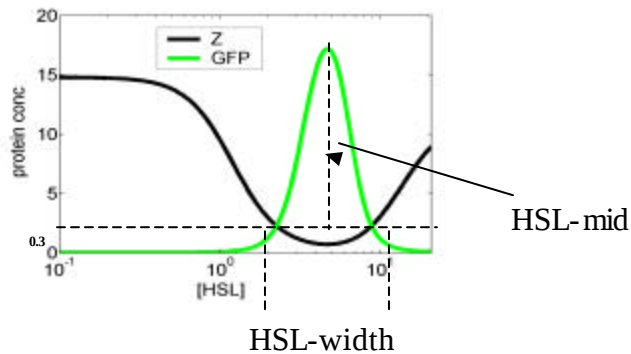




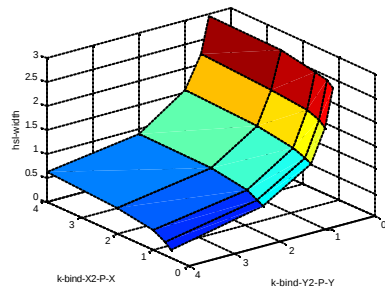


Engineering Circuit Characteristics

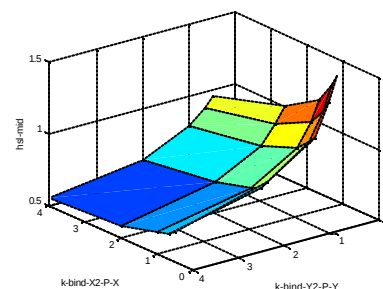
- HSL-mid: the midpoint where GFP has the highest concentration
- HSL-width: the range where GFP is above $0.3\mu\text{M}$



Tuning the Range: Repressor/Operator Affinities



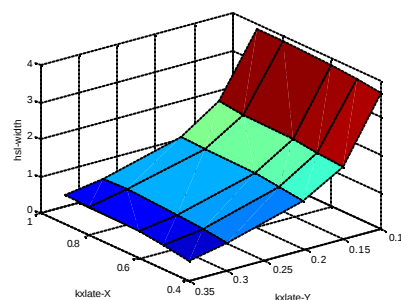
range width
versus
X & Y repressor efficiencies



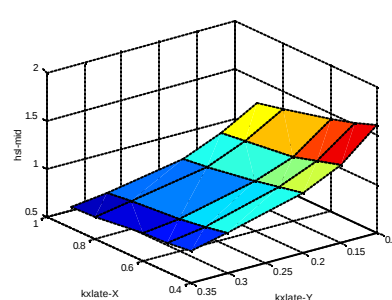
range mid-point
versus
X & Y repressor efficiencies

rep/op affinity increases → transfer-curve shifts left

Tuning the Range: Ribosome Binding Sites



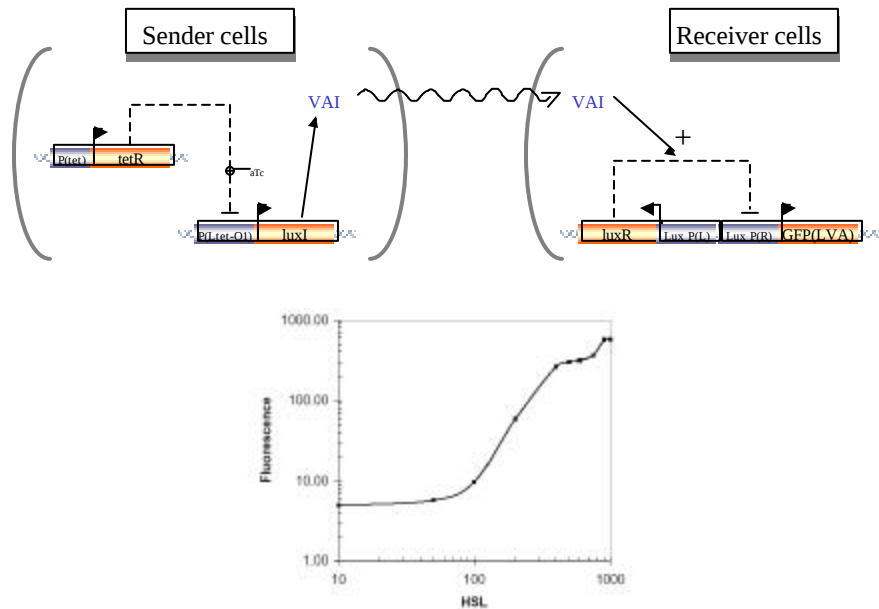
range width
versus
X & Y RBS efficiencies



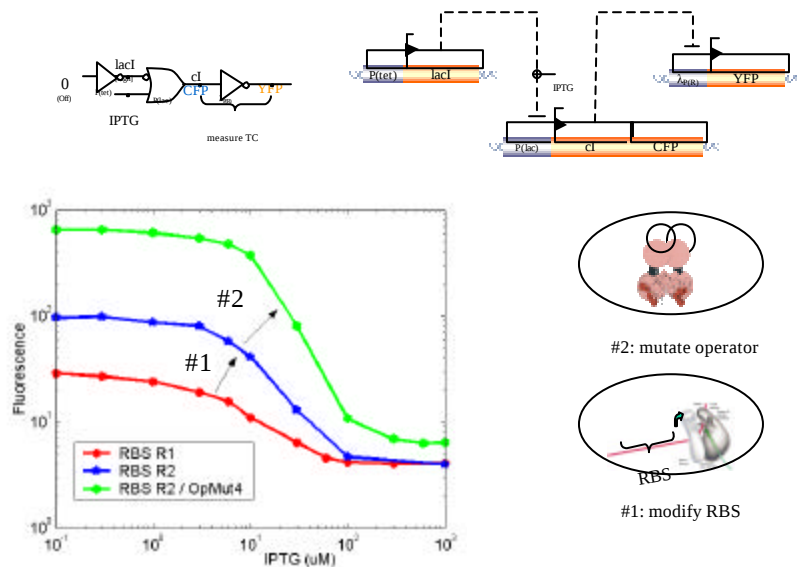
range mid-point
versus
X & Y RBS efficiencies

RBS efficiency increases → transfer-curve shifts left

HSL Detection

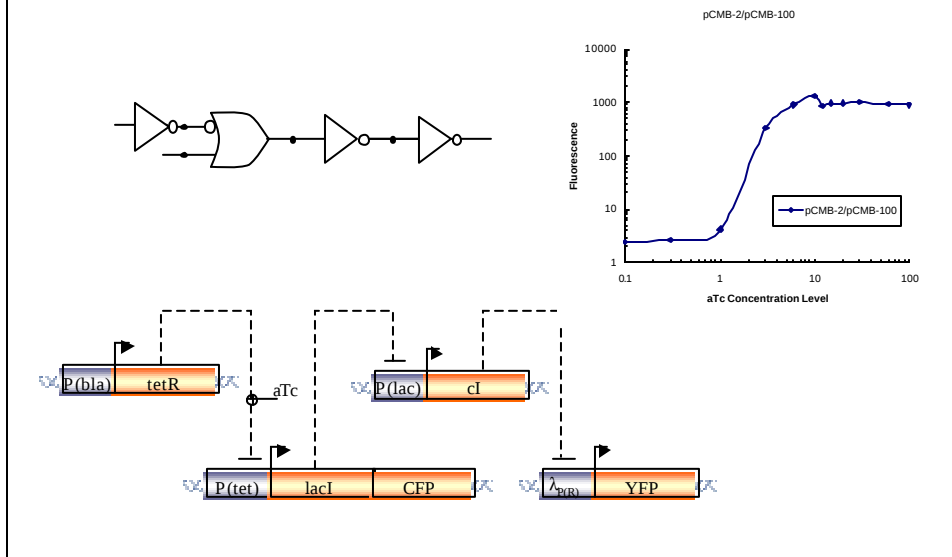


Low Threshold Component



Weiss & Basu, NSC 2002

Genetic Circuit for High Threshold



Circuit Design Principles

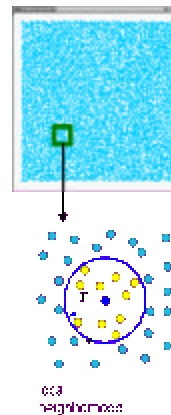
- Separation of low threshold and high threshold
 - RBS efficiency of X must be higher than that of Y
 - Binding affinity of X to its respective promoter has to be higher than that of Y
- Constants associated with Y have more impact on range-width and range-midpoint
 - Y passes through an additional gain stage
- Leakiness and sensitivity of lux promoter determines the lower bound of detection of acyl-HSL

Amorphous Computing

Programming Cell Aggregates

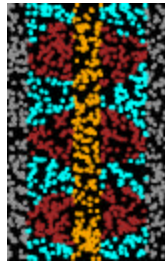
- Amorphous Computing:

“How does one engineer prespecified, coherent behavior from the cooperation of vast numbers of unreliable parts that are interconnected in unknown, irregular, and time-varying ways.”

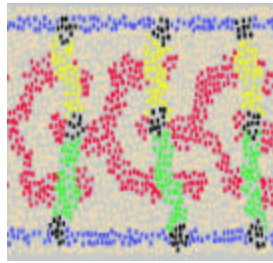


- An aggregate of cells is an example of an amorphous computing substrate

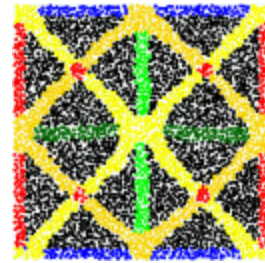
Engineering Coordinated Behavior



MCL
[Weiss, 1998]



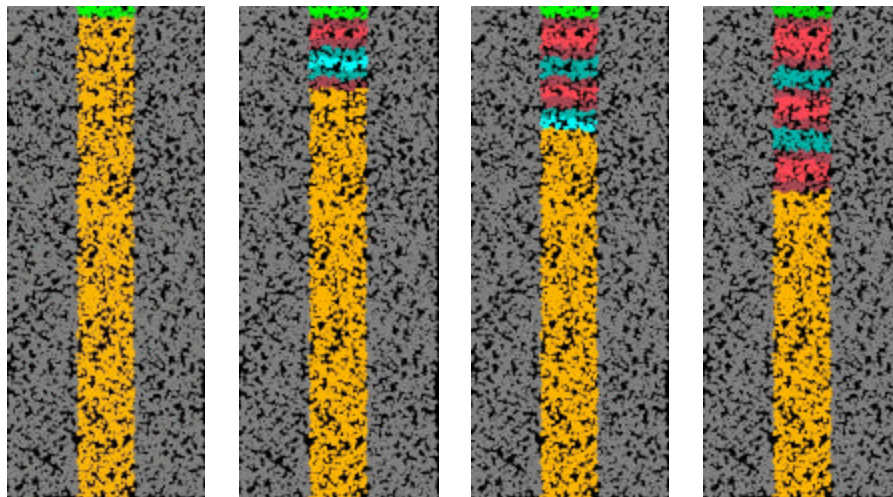
GPL
[Coore, 1997]



Origami
[Nagpal, 2001]

- High-level specifications for pattern formations
- Translate programs to genetic circuits

Another Example: Differentiation



Cells differentiate into bands of alternating C and D type segments.

Microbial Colony Language (MCL)

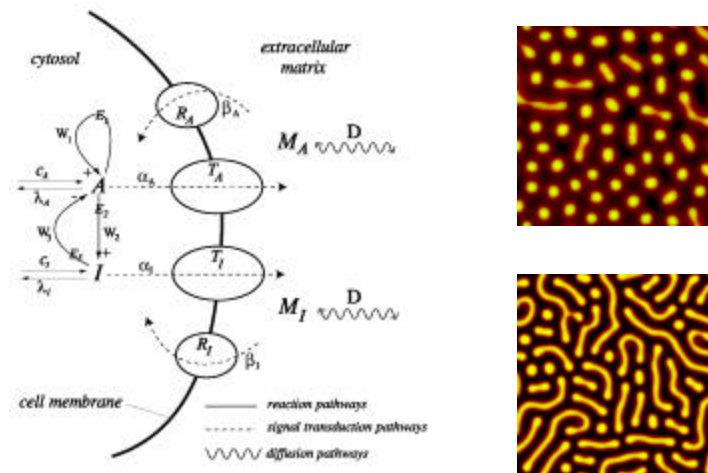
A program for creating segments:

(start	(created	}message
Crest	(br C D)	}condition
((send (make-seg C 1) 3)))	((set Waiting 10)))	}actions
((make-seg seg-type seg-index)	(*	
(and Tube (not C) (not D))	(and Bottom C 1 (Waiting (= 0)))	
((set seg-type)	((send (make-seg D 1) 3)))	
(set seg-index)		
(send created 3)))	(*	
	(and Bottom D 1 (Waiting (= 0)))	
((make-seg) (= 0))	((send (make-seg C 2) 3)))	
Tube		
((set Bottom)))	(*	
	(and Bottom C 2 (Waiting (= 0)))	
((make-seg) (> 0))	((send (make-seg D 2) 3)))	
Tube		
((unset Bottom)))	(*	
	(and Bottom D 2 (Waiting (= 0)))	
	((send (make-seg C 3) 3)))	

The Microbial Colony Language

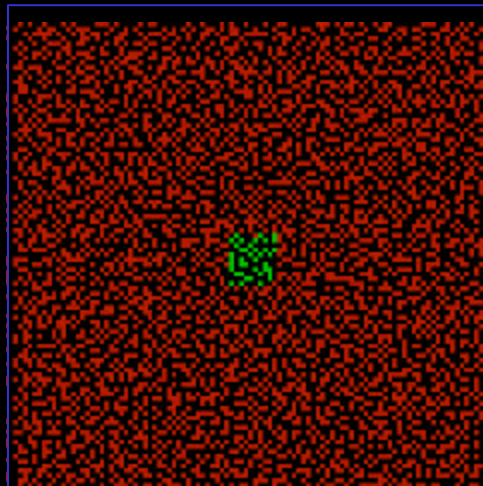
- Language primitives:
 - asynchronous rules
 - boolean state variables
 - boolean logic
 - local communications with chemical diffusion
- These primitives can be mapped to engineered biochemical processes

Reaction/Diffusion Pattern Formation [Millonas/Rauch]

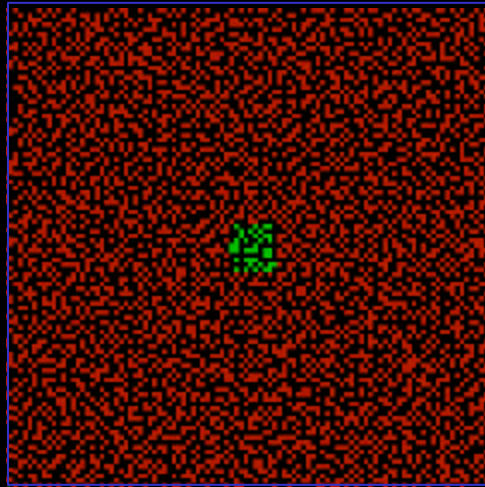


Kinetic rates determine emergence of patterns

Reaction/Diffusion Simulation



Reaction/Diffusion Simulation II



Future Work

- Quantitative prediction of engineered cell behavior
- Self-perfecting genetic circuits
- Intercellular communication architectures
- Signal processing circuits
- Additional CAD tools
- Bio-fab
 - Large scale circuit design, production, and testing
- Simpler & more complex organisms:
 - Eukaryotes
 - Mycoplasmas
- Biologically inspired logic gates
- Molecular scale fabrication



vs.

